Identification of Lipase Producers*Staphylococcus aureus* and Studyingthe Effects of Some Physicochemical Factors on Lipase Production

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ABSTRACT

Lipases catalyze both the hydrolysis and synthesis of triacylglycerols. In addition to the industrial uses of lipases, there is an evolving literature on their role as important microbial virulence factors. One hundred samples were collected from different body sites and lesions of in and out patients from both sexes who attended Al-Kadhimyia Teaching Hospital and Medical Laboratories in Baghdad Teaching Hospital during the period from March, 2012 until September, 2012 for the isolation and identification of *Staphylococcus aureus*. Identification of bacterial isolates was performed according to the standard protocols. According to lipase activity measurement, it was revealed that isolate no. 12 was the higher producer as compared to the others and thus it was selected for further work. The results demonstrated that the best lipase production was in the alkaline pH (8) and at temperature of 30°C. It is also shown that the addition of 0.001M BaCl₂ and 2mM KCl to the culture media has dramatically increased lipase activity. Conversely, the addition of 10mM NaCl, 0.001M FeSO₄.7H₂O, and 2M NaF has caused significant reduction in lipase activity.

تشخيص عزلات المكورات الذهبيه العنقوديه المنتجه لانزيم اللايبيز ودراسة تاثير بعض العوامل الفيزيوكيمياويه على انتاج اللايبيز

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الخلاصة

يحفز انزيم اللايبيز عملية التحلل المائي والتخليق لمركبات ثلاثية الاسل الكليسورليه وبالاضافة الى الاستعمالات الصناعية لللايبيزات فان هنالك العديد من المصادر التي تشير الى اهميتها كعوامل ضراوه مهمه للمكروبات. تم جمع مئه عينه من اماكن وأفات مختلفه من جسم الانسان للمرضى من كلا الجنسين الذين زاروا العياده الخارجيه او كانوا راقدين في مستشفى الكاظميه وأفات مختلفه من جسم الانسان للمرضى من كلا الجنسين الذين زاروا العياده الخارجيه او كانوا راقدين في مستشفى الكاظميه وبغداد التعليميين للفتره من اذار لسنة 2012 ولغاية ايلول من نفس السنه. وكان الغرض من جمع هذه العينات هو لعزل وتشخيص بكتريا المكورات العنقوديه الذهبيه تم تشخيص العزلات الاولي بالطرق الكيموحيويه وبالاعتماد على الطرق القياسيه وتشخيص بكتريا المكورات العنقوديه الذهبيه تم تشخيص العزلات الاولي بالطرق الكيموحيويه وبالاعتماد على الطرق القياسيه وتشخيص بكتريا المكورات العنقوديه الذهبيه تم تشخيص العزلات الاولي بالطرق الكيموحيويه وبالاعتماد على الطرق القياسيه وتشخيص بكتريا المكورات العنوديه الذهبيه تم تشخيص العزلات الاولي بالطرق الكيموحيويه وبالاعتماد على الطرق القياسيه وبني في والتخلي ولينات هو لعزل وبنياس فعالية انزيم اللايبيز تبين ان العزله رقم 12 هي الاعلى في انتاجية هذا الانزيم مقارنة بالعزلات الاخرى ولذلك تم اختيارها لاجراء باقي التجارب. . اظهرت نتائج الدراسه ان افضل انتاج لانزيم اللايبيز كان في البيئه القاعديه (pH 8) وعند درجة حراره 30 مؤيه كما تبين ان المافة 0.001 ملي مولر من ثنائي كلوريد الباريوم و 2 ملي مولر من كلوريد البوتاسيوم دريا وي دياد من كلوريد الموديون ولي معاري الكيبيز مالي درياده ملحوظه في فعالية انزيم اللايبيز على مولر من شنائي كلوريد الباريوم و 2 ملي مولر من كلوريد البيبين الى من كلوريد الموديوم و 0.000 ملي مولر من وليبين الى المائي ولي مالوريد المودي منان مالايبينه القاعديه (0.00 ملي مولر من كلوريد الموديوم و 0.000 ملي مود من مالي ماليبيز على ماليبيز مال مريوم و 3 ملي مولر من كلوريد الموديوم و 0.000 ملي مولر من حامض الكبيني من مامول الكيبيز على المانيبي الايبيز مالي مالي مالي ماليبين مالي ماليبيوم و 10 مالي مالي ماليبي ماليبيوم مو 4 مالي ماليبيو مالي ماليبين ماليبيو مالي مالي مالي مالي مالي ماليبيبي مالي ماليبيو ماليما ماليبيو ماليما مالي ماليما مالي

1. INTRODUCTION

Staphylococcus aureus is a ubiquitous bacterium that is generating increasingly bad press coverage due to its propensity to adopt a pathogenic lifestyle in hospital and community settings.^[1]Lipases catalyze both the hydrolysis and synthesis of triacylglycerols. In addition to the industrial uses of lipases, there is an evolving literature on their role as important microbial virulence factors. ^[2] *S. aureus* colonies are found in approximately 30% of the general population ^[3] and causes pyogenic infection in man and is the common cause of boils, carbuncles, impetigo and infection of surgical or accidental wounds and burns.^[4] Various forms of trauma resulting around superficial abscess such as squeezing a boil may force large number of staphylococci into the blood from where they are carried into many tissues of the body; thus, systemic infections and abscesses may be produced in various deep organs such as liver, lung or brain tissues.^[5]

The putative roles of microbial extracellular lipases include digestion of lipids for nutrient acquisition, adhesion to host cells and host tissues, synergistic interactions with other enzymes, nonspecific hydrolysis due to additional phospholipolyticactivities, initiation of inflammatory processes by affecting immune cells, and self-defense mediated by lysing competing microflora.^[6]

The aims of the study is to identify lipase producer *S. aureus* and study the effects of pH, temperature, and addition of inorganic ions on lipase production; for these parameters to further manipulate the produced lipase for eventual practical applications.

2. MATERIALS AND METHODS

One hundred samples were collected from different body sites and lesions of in and out patients from both sexes who attended Al-Kadhimyia Teaching Hospital and Medical Laboratories in Baghdad Teaching Hospital during the period from March, 2012 until September, 2012 for the isolation and identification of *S. aureus* which was identified through microscopic detection, cultural examination, and biochemical tests.API Staph system was applied for confirming the primary bacterial identification.The method described by Bier^[7] for the measurement of lipase activity through the titration of resulted fatty acids with 0.05 N sodium hydroxide was applied.Solution of reaction was prepared according to Aisakaand Terada.^[8]Enzyme activity was calculated according to Varley and Bell^[9].Isolate No. 12 showed the higher lipase activity among all other isolate thus it was selected as a candidate for further work.

Ten screw capped universals were prepared with 10 ml of brain heart infusion broth inside each, then the pH of the media in each universal was adjusted to 4.5, 5, 5.5, 6, 6.5, 7, 7.5, 8, 8.5 and 9. Each screw capped tube was inoculated with *S. aureus* isolate no. 12 and incubated for 24 hrs. Afterwards tubes were collected and enzyme activity was measured for each tube separately. To determine the optimum temperature for lipase production, six screw capped tube each loaded with ten milliliters of brain heart infusion broth were inoculated with lipase producer *S. aureus* isolate no. 12and were incubated with different degrees of temperature (25, 30, 35, 40, 45, and 50°C) for 24 hrs. ^[10]

Another six screw capped tubes were prepared to have ten ml brain heart infusion broth each; tubes one through five contained 10mMNaCl, 0.001M BaCl₂, 2mMKCl, 0.001M FeSo₄.7H₂O, and 2M Naf, respectively. Tube number six contained only the culture media and

left untreated as control. The tubes were inoculated with lipase producer *S. aureus* isolate no. 12 and were incubated for 24 hrs. Enzyme activity was measured for each tube separately.^[10]

3. RESULTS

From a total of 100 specimens enrolled in the current study, *S. aureus* constituted 50 % of all specimens and it was isolated as a common pathogen that can cause serious infections in various body sites and tissues (Table 1).

| Cases | No. of isolates | % |
|--------------------|-----------------|------|
| Wounds swabs | 15 | 30% |
| Ear swabs | 5 | 10% |
| Sputum | 10 | 20% |
| Urine samples | 6 | 12% |
| Urethral discharge | 4 | 8% |
| Vaginal swabs | 3 | 6% |
| Blood samples | 7 | 14% |
| Total | 50 | 100% |

Table 1:Ranking of *S. aureus* and the percentage of each infection from a total of 50 cases.

Lipase activity was assayed for all the fifty isolates enrolled in the current study and it appeared that isolate number 12 showed the maximum enzyme activity (600 \square mole/ml) as compared to all other isolates (Figure 1).

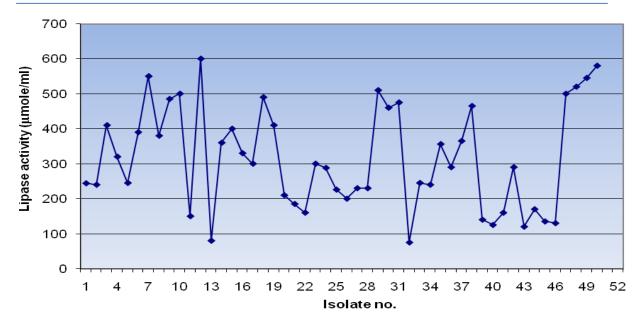


Figure 1: Isolates of S. aureus with different lipase activities.

The results indicated that lipase production was maximal at pH near alkalinity (8) as indicated by an enzyme activity of 540 \Box mol/ml and it was also clear that lipase production was significantly reduced as the pH of the medium shifts gradually towards acidity but showed little effect when pH progressed towards alkalinity (Figure2).

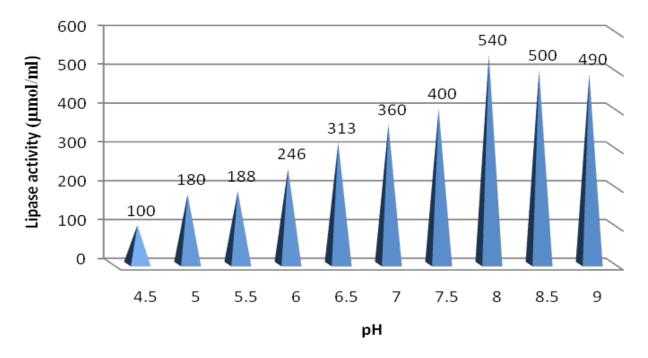


Figure2: Effect of the pH on production of the lipase from the local isolate of S. aureus (12).

A range of incubation temperatures was applied to specify the optimum temperature of incubation. The results showed that the maximum enzyme activity (1115 \square mole/ml) was obtained when incubation temperature was 30°Cfor a period of 24 hrs (Figure 3).

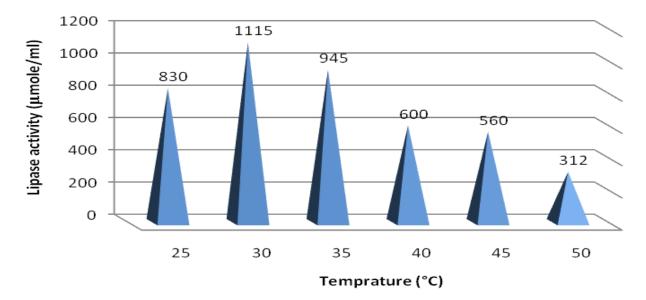


Figure3:Effect of temperature on the production of lipase from a local isolate of S. aureus (12).

Five inorganic ions were selected to evaluate their effects on lipase activity. It was shown that the addition of BaCl₂ and KCl had dramatically increased lipase activity, while the addition of NaCl, H₂SO₄.7H₂O, and Naf had negative influence on lipase activity as compared to the control (Figure 4).

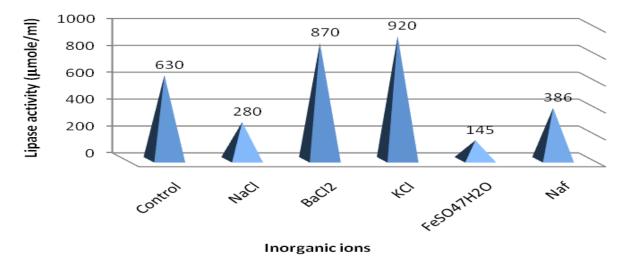


Figure 4: Effect of inorganic ions on the production of lipase from a local isolate of *S. aureus* (12).

4. DISCUSSION

Staphylococci remain among the most important pathogens isolated in the clinical microbiology laboratory ^[11]. *S. aureus* was superabundant and easy to collect from different body sites and lesions because of the following reasons: it's a leading cause of hospital-acquired infections and it is the primary cause of lower respiratory tract infections, pneumonia surgical site and cardiovascular infections^[12,13] and the second leading cause of nosocomial bacteremia ^[14]. Infections with *S. aureus* are especially difficult to treat because of evolved resistance to antimicrobial drugs ^[15].

Data presented in Table 1 showed that *S. aureus* consisted of 50% of all enrolled isolates. Similar results were obtained by other workerswho reported percentages that ranged between 27.6% to 67.3% indicating that this bacterium was a dominant pathogen.^[16, 17]The differences in the frequency of isolation could be attributed to factors such as the anatomical site of infection, type of lesion, age, and geographical area of isolation.^[18]

Our results showed that the maximum enzyme activity was 600 \square mole/ ml. The ability of microorganisms to produce enzyme is influenced by environmental conditions such as temperature, pH and presence of inducers or repressors.^[19] The results also indicated that lipase production was maximal when the pH of the medium was alkaline (pH 8).In agreement with the results of the current study, an alkaline pH 8.5 was optimal for maximum lipase activity produced by *P. fluorescens* in medium which contained emulsified olive oil as the carbon source,^[20] while the study of Sztajer*et al.*,^[21] contradicted the results of the present study when it was found that maximal production of extracellular lipase from *P. Citiinum* was at pH 7.2. Interestingly, researchers working with lipase from *Aspergillusoryzae* recorded higher enzyme activity when the medium approached prominent acidity of pH 4.0 ^[22].

Bacterial lipases having neutral ^[23], basic^[24] and acidic^[25] optimum pH have been reported by different workers. It was found that bacterial lipases show different degrees of enzyme activity in relation to the type of enzyme produced by the microorganism and it was also stated that the optimum pH for bacterial growth and enzyme activity is not necessary match. The pH of the medium can affect the ionizing groups found in the enzyme substrate complex and this will lead to decreased saturation of the enzyme active site with the substrate or a decrease in enzyme stability or both resulting in altered enzyme stability under changing degrees of pH ^[26].

The results of the current study revealed that the activity of the enzyme peaked at temperature 30° C, and it also revealed that the higher temperature the greater the drop in the activity of the enzyme, while the drop in the activity of the enzyme was less when the condition of incubation directed towards colder degrees. This might be explained due to limited adaptation of the enzyme to a reasonable increase in temperature. Many studies were held to detect the optimum temperature for lipase production and some of these results were similar to the results of the present study. Our results are similar to that obtained by Dong *et al.*, ^[27]who were working with *pseudomonas* and stating that optimum lipase activity attained at 30°C for 72 hrs of incubation. In a previous study by Sidhu *et al.*, ^[28] an extra cellar *Bacillus* lipase had optimum activity at 50 °C, while Wang *et al.*, ^[29] showed that the maximum enzyme activityfrom *Bacillus* strain was achieved at 60°C. The results of the present study contradict that of Tyski *et al.*, ^[30]

The marked decrease in enzyme activity at decreased temperature of incubation can be explained by the fact that under low temperatures bacterial growth is suboptimal and the biosynthesis of enzymes is at its lowest level, while normal enzyme production is restored at temperatures close to the forties due to the ability of bacteria to grow well around these degrees.^[31]

In respect to the selected inorganic ions, it was shown that addition of KCl and BaCl₂ had increased lipase activity, while addition of NaCl,H₂SO₄.7H₂O and Naf had negative influence on lipase activity. Chartrain *et al.*,^[32] observed that an extracellular lipase of *P.aeruginosa* was stimulated by CaCl₂, while the activity was not affected by Mg ⁺², Mn⁺², Na⁺ and K⁺. It was proposed that calcium is needed as a cofactor for catalytic activity.^[33]

The inhibitory effect of certain ions on lipase activity of *P. fragi* was observed by Nashif and Nelson ^[34]. Also, it had been noticed that some ions (Zn⁺⁺ and Hg⁺⁺) hadan inhibitory effect on lipase activity.^[35]It was observed that SrCL₂, BaCl₂, and MgCl₂ were the best activators of lipase and this finding was explained by the absolute requirement shown by most lipases to fatty acids acceptors (divalent cations)^[36]. In a marked agreement with the results of the present study, a researcher also showed that lipase activity was enhanced by the addition of KCl to the culture medium of staphylococci. The stimulatory effects of salts can be explained by the fact that they can facilitate or interfere in the interaction between active binding site of the enzyme and the substrate, while the inhibitory effects of salts on enzyme activity may be due to the changes in the ionic strength of the environment which can affect the ionic state of both the enzyme and substrate.^[7]In another study that agreed with the results of the present one, three forms have been purified and characterized and found to have their activity strongly inhibited by Ag^{+2} and Hg^{+2} , whereas Ca⁺² and Mg⁺² enhance lipase activity.^[37]In Algerian study conducted at Sidi bel abbes on samples collected from waste water it was mentioned that the activity of lipase produced by P. aeruginosa was enhanced by Ca⁺² and Mg⁺² but strongly inhibited by heavy metals Zn⁺², Cu⁺² and $Mn^{+2[38]}$ which is in agreement with the results of the present study.

CONCLUSIONS

S. aureus was identified as a major pathogen affecting multiple body organs and tissues and can cause considerable health problems and that lipase production was prominent among isolates belong to this species. It is concluded that lipase production was affected by changing some physicochemical parameters.

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